

Food plant diversity as broad-scale determinant of avian frugivore richness

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The causes of variation in animal species richness at large spatial scales are intensively debated. Here, we examine whether the diversity of food plants, contemporary climate and energy, or habitat heterogeneity determine species richness patterns of avian frugivores across sub-Saharan Africa. Path models indicate that species richness of *Ficus* (their fruits being one of the major food resources for frugivores in the tropics) has the strongest direct effect on richness of avian frugivores, whereas the influences of variables related to water–energy and habitat heterogeneity are mainly indirect. The importance of *Ficus* richness for richness of avian frugivores diminishes with decreasing specialization of birds on fruit eating, but is retained when accounting for spatial autocorrelation. We suggest that a positive relationship between food plant and frugivore species richness could result from niche assembly mechanisms (e.g. coevolutionary adaptations to fruit size, fruit colour or vertical stratification of fruit presentation) or, alternatively, from stochastic speciation–extinction processes. In any case, the close relationship between species richness of *Ficus* and avian frugivores suggests that figs are keystone resources for animal consumers, even at continental scales.

Keywords: Africa; coevolution; community assembly; macroecology; plant–frugivore interactions; spatial autoregressive model

1. INTRODUCTION

A large number of hypotheses have been proposed to explain patterns of species richness at broad spatial scales (Willig *et al.* 2003). Based on high correlations with species richness, contemporary climate and energy variables (e.g. precipitation, temperature and/or evapotranspiration) are often thought to explain spatial variation in species richness better than any other non-climatic variable (Wright 1983; Hawkins *et al.* 2003a; Currie *et al.* 2004). However, a number of other factors also determine broad-scale patterns of species richness, including topography, habitat diversity, or regional and evolutionary history (e.g. Rahbek & Graves 2001; Jetz & Rahbek 2002; Willig *et al.* 2003). Despite a century of debate about the primary determinants of species richness, the underlying causal mechanisms behind the patterns still remain vague (Willig *et al.* 2003; Currie *et al.* 2004; Rahbek *et al.* 2007).

For vascular plants, it is widely argued that precipitation and ambient energy are the main drivers of species richness (Hawkins *et al.* 2003a; Field *et al.* 2005). Water availability, heat and light directly influence plant growth and productivity and are essential to plant physiological processes (Waide *et al.* 1999; Field *et al.* 2005). Higher productivity might result in more species because physiological tolerances of individual species vary for different climatic conditions ('physiological tolerance hypothesis'; Currie *et al.* 2004), or, alternatively, because

more productive areas are warmer and evolutionary rates might be faster at higher ambient temperatures ('speciation rate hypothesis'; Allen *et al.* 2006). For animals, especially for endotherms, the relationships between species richness and water, energy and climate are less pronounced than for plants (Rahbek & Graves 2001; Jetz & Rahbek 2002; Hawkins *et al.* 2003a,b). One likely explanation is that energy might not directly influence animal species richness via its effect on animals' physiological requirements or evolutionary rates, but rather indirectly via trophic relationships (Wright 1983; Hawkins *et al.* 2003a,b; Currie *et al.* 2004). This hypothesis assumes that richness of animals is determined by the abundance, distribution and diversity of food resources (e.g. plant biomass for herbivores, fruits for frugivores).

At small spatial scales, animal species richness can be associated with the abundance, diversity or partitioning of food resources (e.g. Herrera 1985; Siemann *et al.* 1998; Novotny *et al.* 2006). This relationship is however difficult to test at large spatial extents because it is difficult to map food resources for animal groups at continental scales (e.g. insects for insectivorous birds). One possibility to test for a link between animal species richness and resources is to relate the species richness of animals to that of their food items (e.g. food plants; Hawkins & Porter 2003; Márquez *et al.* 2004; Novotny *et al.* 2006). However, correlations between animal and plant species richness can also result from both groups responding similarly to the same environmental variables. After accounting for these environmental variables, a convincing dependency of animal on plant species

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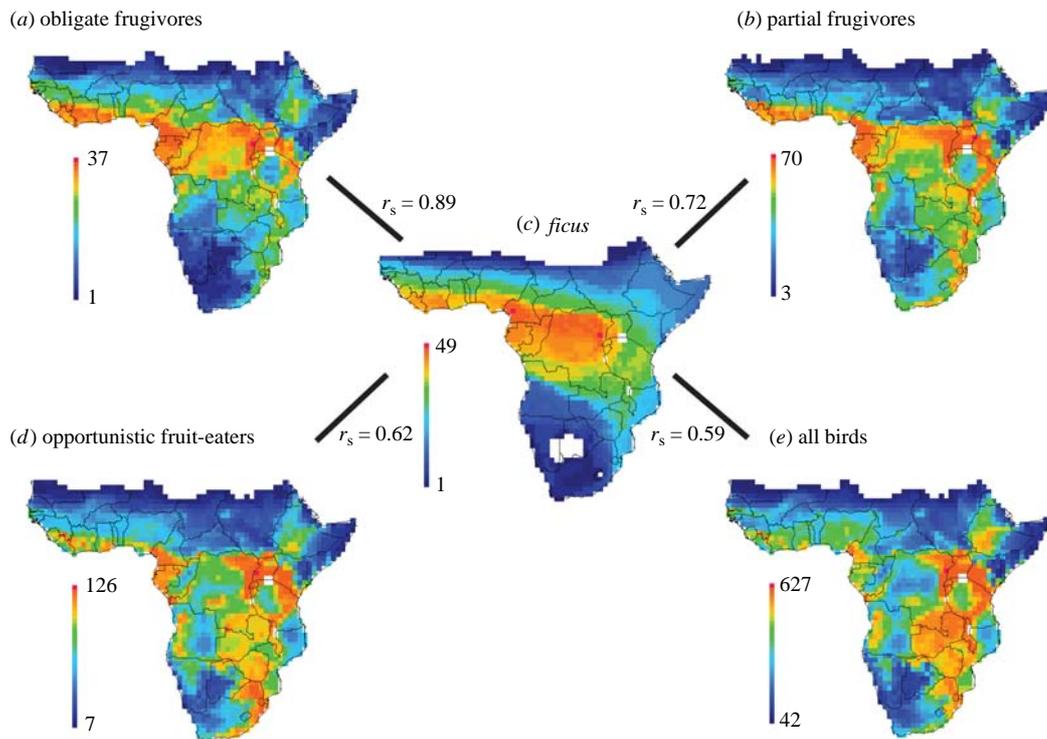


Figure 1. Geographical patterns of species richness in sub-Saharan Africa. (a) Obligate frugivores (92 species), (b) partial frugivores (200 species), (c) all *Ficus* trees (86 species), (d) opportunistic fruit eaters (290 species) and (e) all breeding birds (1771 species). Equal frequency classification is shown, with colour ramps indicating minimum (dark blue, bottom of legend) and maximum (dark red, top of legend) species richness. Note that the scale of richness differs among figures.

richness has not been demonstrated so far at broad spatial scales (Hawkins & Porter 2003; Hawkins & Pausas 2004; Márquez *et al.* 2004).

Plant–frugivore interactions might be an ideal model system for continental analyses of animal and plant species richness. Most frugivorous animals heavily rely on fruits, particularly in the tropics (Fleming *et al.* 1987). In a number of fine-scale field studies, it has been shown that the richness of frugivorous animals is largely dependent on fruit availability (e.g. Herrera 1985; Fleming *et al.* 1987; Bleher *et al.* 2003). Among the fruiting plants, the fig genus (*Ficus*) has been considered to be a keystone plant resource for many frugivores owing to large crop sizes and asynchronous fruiting patterns throughout the year (Terborgh 1986; Lambert & Marshall 1991; Shanahan *et al.* 2001a; Bleher *et al.* 2003; Harrison 2005; but see Gautier-Hion & Michaloud 1989). Thus, the diversity and abundance of figs might set the carrying capacity for frugivorous animals in the tropics. Correspondingly, Goodman & Ganzhorn (1997) proposed that avian frugivore richness might depend directly on species richness of *Ficus* trees. However, no rigorous test of this ‘fig–frugivore–richness hypothesis’ has been conducted at a large regional scale such as a continent.

In this study, we examine whether the richness of *Ficus* species at a continental scale (i.e. sub-Saharan Africa) influences avian consumer richness by examining a comprehensive database with a resolution of 1° latitude and longitude, summarizing the distribution of all breeding birds ($n=1771$), all *Ficus* species ($n=86$) and five climatic and environmental variables (precipitation, temperature, productivity, topography and ecosystem diversity). We classify frugivorous birds into three classes (obligate, partial and opportunistic fruits eaters) and

predict the association between frugivore and *Ficus* richness to be stronger for those frugivores that are more specialized on fruit eating. We apply path analysis to disentangle inter-correlations between variables and compare the results of this non-spatial method with those of spatial regression models that account for the spatial autocorrelation structure within our dataset.

2. MATERIAL AND METHODS

(a) Bird data

We used an updated version (29 September 2005) of the comprehensive distribution database of African breeding birds compiled by the Zoological Museum, University of Copenhagen (see Burgess *et al.* (1998) and Brooks *et al.* (2001) for methodology; Jetz & Rahbek (2002) for sources used for mapping). Maps for each species represent a conservative extent-of-occurrence extrapolation of the breeding range at a resolution of $1^\circ \times 1^\circ$ cells (latitude–longitude). Data were compiled from the standard reference works and dozens of other published references (including recent atlases and unpublished research) and, for difficult regions and taxa, experts’ opinions were sought (the full list of sources is available at http://www.zmuc.dk/commonweb/research/bio_data.htm). Most of the northern part of continental Africa, the Sahara, is marked by extreme species scarcity (Jetz & Rahbek 2002) and almost all species in it and North of it belong to the Eurasian biome. We thus focused our analyses on all 1771 breeding bird species south of the Saharan desert ecoregion (figure 1e) with ecoregion boundaries for the South Sahara as northern boundary (Olson *et al.* 2001). Our sub-Saharan database contains 434 789 records on 1737 cells. The extent of the grid was chosen to be similar to the one used by Jetz & Rahbek (2002) to make the results comparable. We therefore excluded cells containing less

Table 1. Predictor variables used to explain spatial variation in richness of avian frugivore species across sub-Saharan Africa.

mnemonic	predictor variables (units)	hypothesis (reference)
FigRich	number of <i>Ficus</i> species per 1° cell (count)	food plant diversity (Goodman & Ganzhorn 1997; Bleher <i>et al.</i> 2003)
Prec	mean annual precipitation (mm yr ⁻¹)	water availability (Rahbek & Graves 2001; Jetz & Rahbek 2002; Field <i>et al.</i> 2005; Hawkins <i>et al.</i> 2003a)
MaxTemp	mean daily maximum temperature (°C)	ambient energy (Jetz & Rahbek 2002; Hawkins <i>et al.</i> 2003a)
NPP	net primary productivity (t C ha ⁻¹ yr ⁻¹)	productivity (Waide <i>et al.</i> 1999; Jetz & Rahbek 2002; Hawkins <i>et al.</i> 2003a)
AltRange	topographic relief (altitudinal range in m)	topographic heterogeneity (Rahbek & Graves 2001; Jetz & Rahbek 2002)
EcoDiv	number of ecosystems in cell (count)	ecosystem diversity (Rahbek & Graves 2001)

than 50% dry land. Cell size varies only slightly with latitude, ranging from 10 188 to 12 308 km². The Worldmap computer program, v. 4.20.24 (1999, P. H. Williams, Natural History Museum, London) was used to overlay the distributional data.

(b) Frugivore classification

The diets of all bird species in our sub-Saharan database were determined from a comprehensive literature survey (see electronic supplementary material S1 for references and classification procedure). We classified all species into three frugivore guilds depending on diet preference for fruits: (i) obligate frugivores (species that primarily feed on fruits, i.e. the only major food items are fruits), (ii) partial frugivores (species that have, beside fruits, other major food items, e.g. terrestrial invertebrates), and (iii) opportunistic fruit eaters (species that only occasionally eat fruits as supplementary food). The three frugivore guilds were characterized by the degree of avian specialization on fruits, with obligate frugivores being most dependent and opportunistic fruit eaters being least dependent on the availability of fruits. Our classification of frugivorous bird species integrates the best knowledge currently available on the feeding behaviour of African birds (see electronic supplementary material S1). For the interested reader, we also provide lists of species of all African frugivores (see electronic supplementary material S2).

(c) *Ficus* data

Individual distribution maps for all *Ficus* species were provided by the Iziko Museums of Cape Town (2005, S. van Noort and J.-Y. Rasplus, available at http://www.figweb.org/Ficus/Species_index/afrotropical_species.htm). The maps are based on country records and the extent of species occurrence is approximated based on habitat affiliations of each species (S. van Noort 2005, Iziko Museums of Cape Town, personal communication). To create a *Ficus* richness map for sub-Saharan Africa, we first georeferenced the maps of each species and digitized the geographical ranges. The ranges of all individual *Ficus* species were then overlaid on a 1° × 1° grid cell map. For each species, we assigned the value 1 indicating species presence for each 1° grid cell when the cell contained more than 10% distribution cover. *Ficus* richness values were then calculated for each cell by adding all presence values. We tested the sensitivity of the 10% distribution cover threshold by calculating *Ficus* richness values from *Ficus* presence maps based on thresholds of 0, 5, 15 and 20% distribution cover. All of the resulting *Ficus* richness patterns were highly correlated with each other (Spearman rank correlations, $r_s > 0.98$) indicating that the arbitrarily chosen threshold of 10% did

not distort the overall *Ficus* richness pattern. Geoprocessing was done with the software ARCVIEW v. 3.2 and ARCGIS v. 9. Taxonomy of *Ficus* follows Berg & Wiebes (1992) and Shanahan *et al.* (2001a; see their Appendix 1). The geographical distributions of different subspecies were pooled as one species. *Ficus thomningii* was used as a synonym for *Ficus petersii* and *Ficus burkei*. A total of 86 *Ficus* species were thus finally distinguished in our study (see electronic supplementary material S3).

(d) Environmental variables

Besides species richness of *Ficus*, we included five environmental variables as potential determinants of the richness pattern of avian frugivores. The environmental variables included two climatic variables related to water input (precipitation) and ambient energy (temperature), a measure of productivity, a measure of topographic heterogeneity and habitat diversity (see table 1 for details). These variables have previously been shown to be strongly correlated with species richness of birds and woody plants at continental scales (Waide *et al.* 1999; Rahbek & Graves 2001; Jetz & Rahbek 2002; Hawkins *et al.* 2003a,b; Field *et al.* 2005). Data for precipitation and temperature were extracted from the mean monthly climatic database for the period 1961–1990 provided by the Intergovernmental Panel on Climate Change (IPCC), available online at http://ipcc-ddc.cru.uea.ac.uk/obs/get_30yr_means.html (see New *et al.* (1999) for methodology). We used mean annual precipitation (mm yr⁻¹) and mean daily maximum temperature (°C) (following Jetz & Rahbek 2002), degraded from 0.5 to 1° resolution. For productivity, we chose net primary productivity (NPP) predictions from the DOLY global model (Woodward *et al.* 1995). Topographic heterogeneity was quantified as altitudinal range (difference between maximum and minimum elevation) of the 1 min digital elevation model presented by Hutchinson *et al.* (1996). Ecosystem diversity was estimated by counting the number of distinct ecosystems in each cell from a recently published map of global ecosystems (Olson 1994; available at <http://edcns17.cr.usgs.gov/glcc/>). While both ecosystem diversity and topographic relief are potentially important predictors in their own right (Rahbek & Graves 2001), they are also rough surrogate variables for habitat heterogeneity.

(e) Statistical analysis

To disentangle the relative roles of predictor variables, many of which covaried (see electronic supplementary material, table S5), and to assess the potential influence of spatial autocorrelation on the robustness of our results, the analysis comprised a three-step process.

In the first step, we calculated Spearman rank correlations (r_s) between all variables in our dataset to examine the strength of the relationships between predictor variables, and between predictor and response variables. In the second step, we applied path analysis (Mitchell 1992; Quinn & Keough 2002), which allows the consideration of hypothesized causal relationships in datasets with more than one dependent variable and effects of dependent variables on one another. Whereas path analysis cannot replace experimental manipulations for detecting causal links between variables, it is one of the few methods to test ecological and evolutionary hypotheses at broad spatial scales (Hawkins & Porter 2003; Márquez *et al.* 2004). Path models are usually presented in path diagrams, where the effect of one variable on another is measured by standardized partial regression coefficients (Mitchell 1992; Quinn & Keough 2002). Path analysis further allows the partitioning of correlation between predictor and response variables (so called 'total effects') into direct and indirect effects. Direct effects are measured by the standardized partial regression coefficients between a predictor variable and a response variable (i.e. the direct link), whereas indirect effects are calculated by adding the products of all standardized partial regression coefficients over all paths between a predictor and a response variable (i.e. including indirect links via other correlated predictor variables; see Mitchell 1992; Quinn & Keough 2002).

Our path models were designed to represent hypotheses of how predictor variables might interact with each other to influence avian frugivore richness, and the links were thus based on *a priori* knowledge or logical relationships among our predictor variables (see references in table 1). Since our main focus was on the potential influence of *Ficus* richness on frugivore richness, we first generated a path model that excluded *Ficus* richness followed by a model to which *Ficus* richness was added. Comparison of the first model with the second model allowed us to evaluate whether *Ficus* richness had a significant effect on frugivore richness itself, or whether it only acted upon frugivore richness through causal relationships with other environmental variables. We assessed the path models using structural equation modelling (SEM), which is an extension of path analysis (see Mitchell (1992) for an introduction). Model evaluation was done by comparing the fitted path models to a baseline model, where observed variables were assumed to be uncorrelated with each other (Arbuckle (2003), see his Appendix C). We used the normed fit index (NFI) as a fit measure (Bentler & Bonett 1980), which ranges between 0 and 1, with values close to 1 indicating a good fit (Arbuckle 2003). The χ^2 goodness of fit test (which is often used to assess the null hypothesis that a path model fits to the data) is invalid in our case, because the large sample size ($n=1737$) would have almost certainly resulted in significant departures from the null hypothesis (Arbuckle 2003, see his Appendix C). We additionally tested whether multiple regression models with all explanatory variables (table 1) explained frugivore species richness better than multiple regression models, where *Ficus* richness was excluded as an explanatory variable. These model comparisons were done with the Akaike Information Criterion (AIC), a model selection criterion which accounts for both model fit and model complexity (Burnham & Anderson 2002).

In our third analysis, we tested for the presence of spatial autocorrelation because our data violate the assumption of independently distributed errors in regression models (Legendre & Legendre 1998), and, as a consequence, the

effects of explanatory variables might thus be exaggerated (Lichstein *et al.* 2002). To quantify the pattern of autocorrelation in our dataset, we calculated Moran's I values (i.e. a measure of autocorrelation) across 20 distance classes (one distance class corresponds to 112 km) and plotted them in so-called correlograms (Legendre & Legendre 1998). We first calculated Moran's I for all raw bird richness data (i.e. obligate frugivores, partial frugivores, opportunistic fruit eaters and all birds), and then fitted multiple regression models with all predictor variables (i.e. models which are equivalent to all direct effects on avian richness in our path models) and recalculated Moran's I on the residuals. Since fitting the multiple regression models with all predictor variables did not remove all of the spatial autocorrelation in our richness variables, we fitted spatial autoregressive models (Cliff & Ord 1981; Cressie 1993) which augment the multiple regression models with an additional term that accounts for patterns in the response variable that are not predicted by explanatory variables, but are instead related to values in neighbouring locations. We then compared the standardized partial regression coefficients (Quinn & Keough 2002) from the spatial autoregressive models to those of our path models (i.e. direct effects on avian richness) to assess whether the relative importance of parameter estimates changes when the spatial autocorrelation structure in our response variables is removed.

All statistical analyses were done with the free software R (R Development Core Team 2005) except for the path models which were calculated with the AMOS software (Arbuckle 2003). The spatial models were calculated as 'spatial simultaneous autoregressive error models' using the R library 'spdep', v. 0.3–25 (2006, R. Bivand, available at <http://cran.r-project.org/src/contrib/Descriptions/spdep.html>). These models are a special type of simultaneous autoregressive models and assume that the response at each location (i) is a function not only of the explanatory variable at i , but also of the values of the response at neighbouring locations (j) as well (Cliff & Ord 1981; Anselin 1988; Cressie 1993). We defined the spatial neighbourhood with a distance of 112 km including the four neighbouring cells that directly join each focal cell (the rook's case). The spatial weights matrix was calculated with a row standardized coding scheme that scales the covariances based on the number of neighbours of each region (see R library 'spdep' for details, reference above). Moran's I values and correlograms were calculated with the R library 'ncf', v. 1.0–9 (2006, O. N. Bjørnstad, available at <http://asi23.ent.psu.edu/onb1/>). To improve the normality of distributions, we transformed all endogenous variables (i.e. those with incoming arrows in the path models) and used transformed values in all regression analyses. Precipitation, maximum temperature and NPP + 1 were log transformed, whereas all richness measures (i.e. species richness of *Ficus*, obligate frugivores, partial frugivores, opportunistic fruit eaters, all birds and ecosystem diversity) were square-root transformed. These transformations yielded the best approximations of normal distributions and were performed to meet the normality of errors assumption (Mitchell 1992; Quinn & Keough 2002). Analyses with untransformed values gave qualitatively similar results.

3. RESULTS

(a) Geographical patterns of species richness

Species richness of obligate avian frugivores ($n=92$) across sub-Saharan Africa is highest in tropical

rainforest regions at equatorial latitudes (figure 1a), particularly, in the coastal areas of West Africa and in the Congo Basin, but also within the East African mountains. Hotspots of obligate frugivore richness are thus not congruent with hotspots of overall bird species richness, which are mainly found in the eastern parts of sub-Saharan Africa (figure 1e). Geographical patterns of species richness of partial frugivores ($n=200$; figure 1b) are more similar to obligate frugivore richness (figure 1a) than to overall bird species richness ($n=1771$; figure 1e), whereas opportunistic fruit eaters ($n=290$; figure 1d) closely resemble overall bird species richness (figure 1e) rather than obligate frugivore richness (figure 1a). The species richness of *Ficus* trees ($n=86$) is highest in the Congo Basin and relatively low in South Africa and along the eastern parts of Africa (figure 1c), and thus largely congruent with obligate frugivore richness patterns (figure 1a).

(b) Determinants of frugivore richness

Simple correlations between *Ficus* and bird species richness indicated that they positively covary across sub-Saharan Africa (figure 1 and see electronic supplementary material, figure S4). As expected, the relationship was strongest for obligate frugivores ($r_s=0.89$), intermediate for partial frugivores ($r_s=0.72$) and lowest for opportunistic fruit eaters ($r_s=0.62$) and overall bird species richness ($r_s=0.59$). Precipitation, NPP, ecosystem diversity and maximum temperature were also strongly correlated ($r_s>0.60$) with avian species richness in almost all cases, and precipitation and NPP highly covaried with each other and with species richness of *Ficus* trees ($r_s>0.84$). Maximum temperature, altitudinal range and ecosystem diversity generally showed weaker correlations ($r_s<0.50$) with other predictor variables (see electronic supplementary material, table S5).

The path model without *Ficus* richness (figure 2a) explained 74.2% of the variance in richness of obligate frugivorous birds, and the measure of fit (NFI=0.891) indicated that the model adequately described the data structure. Precipitation had the strongest direct effect on richness of obligate frugivorous birds followed by NPP, altitudinal range, ecosystem diversity and maximum temperature (figure 2a). Including richness of *Ficus* trees in the path model improved the explanatory power (81.7%) and the overall fit of the model (NFI=0.920), and we thus consider this path model (figure 2b) a better description of obligate frugivore richness patterns. Model selection based on AIC values also indicated that a multiple regression model with *Ficus* species richness data better (i.e. had a lower AIC value) than a multiple regression where *Ficus* richness had been excluded (AIC=2838, Δ AIC=593). In the path model with *Ficus* richness (figure 2b), the direct effect of precipitation on richness of obligate frugivorous birds was very low (0.095) and richness of *Ficus* trees instead became the most important variable with the strongest direct effect (0.546) on richness of obligate frugivorous birds (table 2; figure 2b). When including indirect effects, the relative importance of precipitation increased (table 2), because it was very strongly correlated with NPP and *Ficus* richness (figure 2b). The total effects of other predictor variables were also higher than their direct effects (table 2),

indicating that they indirectly affected frugivore richness via other variables.

Replacing obligate frugivores in our path model (figure 2b) with partial frugivores, opportunistic fruit eaters and all birds resulted in less-explained variance in avian species richness (partial frugivores, 73.7%; opportunistic fruit eaters, 70.3%; all birds, 66.4%) than the original path model (obligate frugivore richness, 81.7%). This trend is consistent with our expectation that the hypothesized causal relationships in the path models should be stronger for birds that are more specialized on fruit eating. Furthermore, these path models showed that direct effects of *Ficus* richness became weaker with decreasing specialization of birds on fruit eating (table 2) which also confirms our expectations. Correspondingly, the AIC values of multiple regression models with all explanatory variables increased with decreasing specialization on fruit eating (AIC_{obligatefrugivores}=2245; AIC_{partialfrugivores}=3452; AIC_{opportunisticfruiteaters}=4275; AIC_{allbirds}=7214).

(c) Effect of spatial autocorrelation

All avian species richness data were spatially autocorrelated over more than 1000 km, although the extent (i.e. distance) differed slightly between frugivore guilds (figure 3). Fitting multiple regression models with all predictor variables (i.e. models with those variables that show direct effects on avian species richness in our path models) reduced spatial autocorrelation in all richness data, indicating that the spatial structure of explanatory variables accounted for some of the spatial autocorrelation structure in the avian richness data. However, our set of explanatory variables could not account for all of the observed spatial structures in our response variables (figure 3). We therefore fitted spatial autoregressive models, which removed almost all of the spatial autocorrelation in richness data across all distance classes (figure 3), indicating that the spatial structure can be explained by including information on the covariance structure from the four neighbouring cells directly joining each focal cell.

The standardized partial regression coefficients of the spatial autoregressive models (table 3) differed from those of the path models (direct effects in table 2), demonstrating that the effects of predictor variables might be exaggerated when using traditional multiple regression or path models. However, despite the changes in parameter estimates, richness of *Ficus* trees still remained the strongest predictor variable to explain the richness pattern of obligate and partial frugivorous birds, respectively. Moreover, its effect still decreased with decreasing specialization of birds on fruit eating (table 3). Besides, the relative importance of other predictor variables to explain obligate frugivore richness did not change when using spatial autoregressive models except precipitation, which became more important in spatial analyses (compare direct effects in tables 2 and 3).

4. DISCUSSION

Our analyses indicate that a positive relationship between species richness patterns of figs (*Ficus* spp.) and avian frugivores exists across sub-Saharan Africa, which suggests that both are linked via resource-consumer

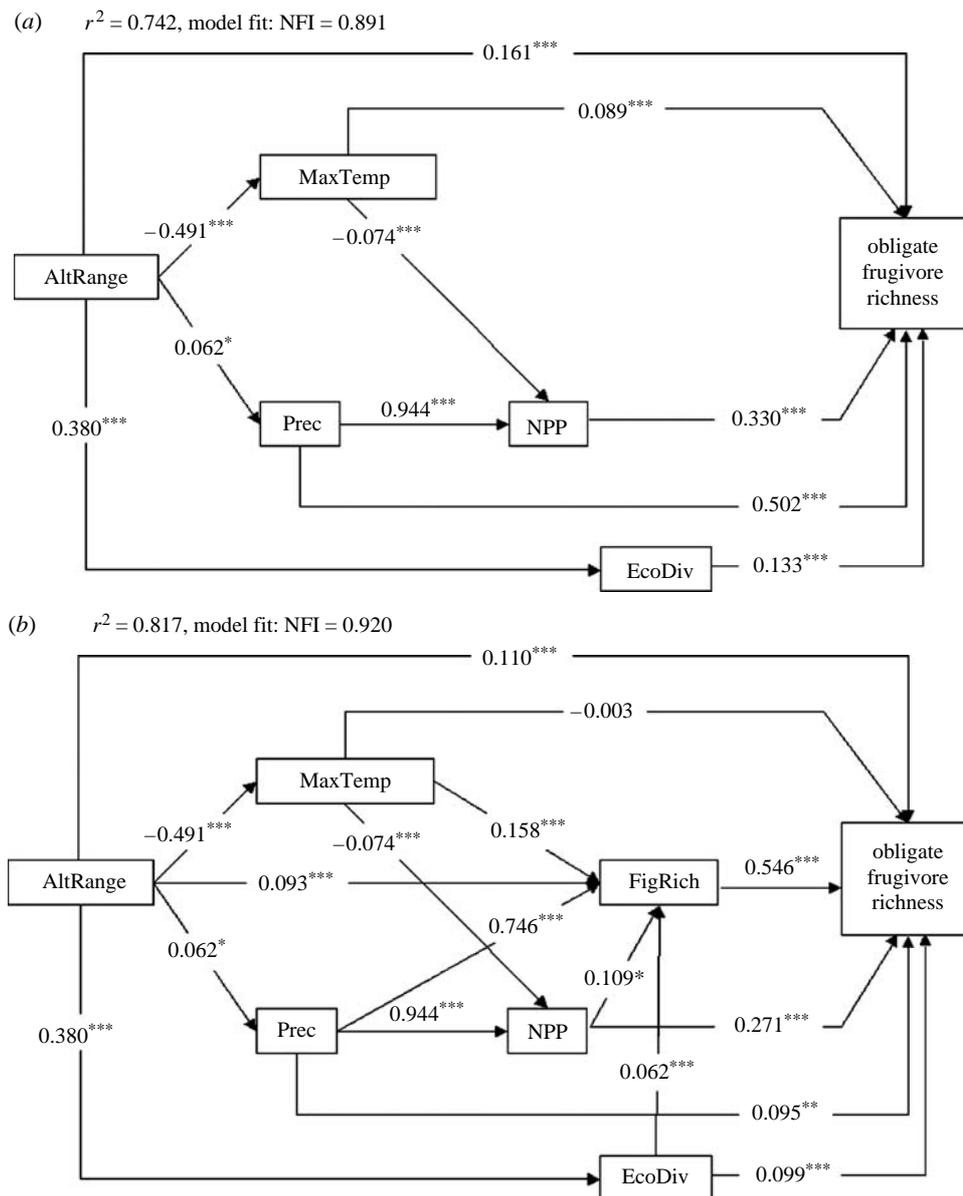


Figure 2. Path models for richness of obligate frugivorous bird species. (a) *Ficus* richness excluded and (b) *Ficus* richness included. Illustrated are direct effects (i.e. standardized partial regression coefficients) and their significance levels (* $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$). r^2 and normed fit index (NFI) are given for each model (see §2 for details).

interactions rather than being caused by similar responses to environmental variables. We thus provide evidence that food plant diversity is an important determinant of avian frugivore richness in tropical regions, even after controlling for confounding environmental variables and spatial autocorrelation. The results also underline the potential role of *Ficus* as a keystone plant resource for avian frugivores in the tropics (Shanahan *et al.* 2001a; Bleher *et al.* 2003; Harrison 2005).

There are a number of mechanisms that could potentially explain a positive relationship between food plant and animal consumer species richness. Some can be based on deterministic processes and niche assembly theory (Graves & Rahbek 2005), whereas others are based on stochastic processes and ecological drift (i.e. neutral theory; Hubbell 2001; see also Colwell *et al.* 2004). One possible explanation for a positive relationship between food plant and frugivore species richness is that a greater number of plant species could potentially provide more

niches for the coexistence of animal species ('niche assembly hypothesis'; Hutchinson 1959). This explanation assumes that animal species specialize on certain food plants or on specific types of resources provided by the plants (Price 2002). For instance, the latitudinal gradient in species richness of herbivorous insects from temperate to tropical regions has been suggested to be a direct function of an increase in plant species richness (Novotny *et al.* 2006). However, this 'reciprocal specialization hypothesis' is unlikely to be relevant for plant–frugivore interactions (Herrera 2002). Most fruit-eating bird species do not specialize on the fruits of a particular plant species. Instead, frugivorous bird species often treat fleshy fruited plant species as interchangeable (Zamora 2000; Herrera 2002). For our study system, we know of only one frugivorous bird species (Bruce's Green-pigeon, *Treron waalia*) that feeds particularly on one single fig species (*Ficus platyphylla*) with the ranges of the two species largely overlapping. Other examples might exist,

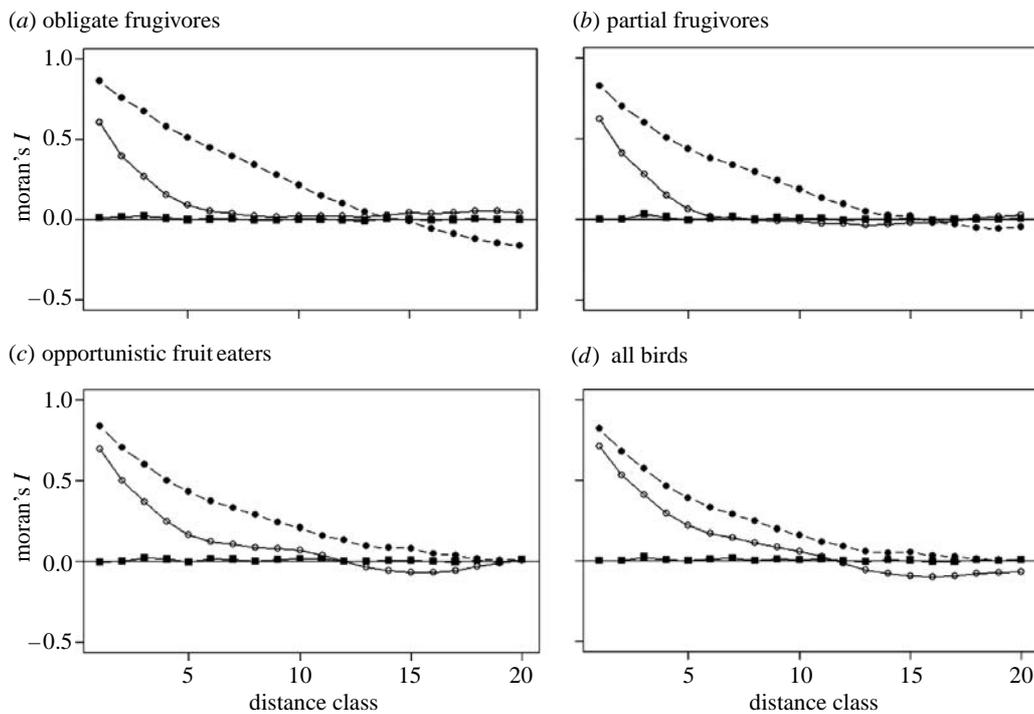


Figure 3. Correlograms for raw data on species richness (solid circles), residuals of multiple regression models (open circles) and residuals of spatial autoregressive error models (solid squares). Both models included all predictor variables (table 1) and species richness of (a) obligate frugivores, (b) partial frugivores, (c) opportunistic fruit eaters and (d) all birds, respectively, as response variables. Multiple regression models thus include all direct effects of predictor variables on avian species richness from our path models. One unit distance class corresponds to 112 km.

Table 2. Standardized direct and total effects of predictor variables on species richness of obligate frugivores (OBL), partial frugivores (PAR), opportunistic fruit eaters (OPP) and all birds (ALL). (Values are derived from path models (figure 2b), which include species richness of *Ficus* trees as predictor variable. Indirect effects are total effects minus direct effects and equal zero if total effects and direct effects have the same values. Mnemonics of predictor variables are explained in table 1.)

predictor variable	direct effects				total effects			
	OBL	PAR	OPP	ALL	OBL	PAR	OPP	ALL
FigRich	0.546	0.454	0.252	0.172	0.546	0.454	0.252	0.172
Prec	0.095	0.018	0.193	0.410	0.814	0.626	0.604	0.611
MaxTemp	0.003	-0.362	-0.421	-0.264	0.064	-0.311	-0.398	-0.243
NPP	0.271	0.236	0.209	0.058	0.330	0.285	0.236	0.077
AltRange	0.110	0.044	0.043	0.137	0.230	0.383	0.388	0.416
EcoDiv	0.099	0.250	0.220	0.267	0.133	0.278	0.235	0.278

but evidence for strong reciprocal specialization between frugivore species and fig or fleshy fruited plant species is generally scarce (Herrera 2002).

Alternatively, a greater number of food plant species could potentially provide more niches for animal consumer species by providing a larger range of resources types. For instance, fruit size is an important attribute of fruits and varies greatly between species (e.g. fruit sizes of *Ficus* species range from 0.5 to 10 cm in diameter; Berg & Wiebes 1992). If frugivores show some specialization on differently sized fruits, then frugivore species richness is likely to increase with a greater range of fruit sizes. There is some evidence for this ‘size-related coupling hypothesis’ (Herrera 2002; Githiru *et al.* 2002; Lord 2004), because fruit size sets limits to fruit ingestion, at least to relatively small-sized birds that swallow whole fruits. It is thus likely that a greater number of *Ficus* species is accompanied with

a larger diversity of fruit sizes (Berg & Wiebes 1992), which may attract a greater size range of fruit-eating birds increasing frugivore species richness (Shanahan *et al.* 2001a). If size-related coupling of fruits and frugivores is the underlying mechanism in our study system, then the correlations between fig and frugivore species richness could result as a by-product of this relationship.

Similar to fruit size, other fruit traits could potentially influence food choice and partitioning of the available fruit spectrum among consumer species (Gautier-Hion *et al.* 1985; Herrera 2002). For instance, frugivorous birds can discriminate among fruits on the basis of colour and might exhibit distinct colour preferences (Herrera 2002). A larger number of *Ficus* species is likely to increase the range of fruit colours (fig colours vary greatly from red, yellow, orange, green, brown to black fruits; Berg & Wiebes 1992), and this might attract a wider range of

Table 3. Standardized partial regression coefficients from spatial autoregressive error models (see §2 for details). All models were calculated as multiple regression models with avian species richness (OBL, obligate frugivores; PAR, partial frugivores; OPP, opportunistic fruit eaters; ALL, all birds) as response variable and all other variables as predictor variables (see table 1 for explanation of mnemonics).

predictor variable	OBL	PAR	OPP	ALL
FigRich	0.382	0.266	0.222	0.231
Prec	0.259	0.253	0.322	0.248
MaxTemp	-0.023	-0.082	-0.032	-0.037
NPP	0.165	0.162	0.160	0.163
AltRange	0.086	0.097	0.070	0.065
EcoDiv	0.077	0.083	0.081	0.123

frugivorous species ('fruit colour-richness hypothesis'). There is some evidence that differences in fruit colour can explain differences in frugivore assemblage structure, at least when considering consumer species across taxa (e.g. when comparing primates and birds; Voigt *et al.* 2004). However, to our knowledge, no study has shown convincingly that certain frugivorous bird species specialize on specific fruit colours. Other fruit traits such as fruit pulp quality (i.e. nutrient composition) could also be critical in food selection of frugivorous animals, but there is generally little evidence that they play an important role in shaping mutual adaptations between fleshy fruited plants and frugivores (Herrera 2002).

Another potential mechanism underlying a positive relationship between fleshy fruited plant and frugivore species richness is that a larger number of food plants are likely to increase the diversity of fruit presentation. For instance, depending on the *Ficus* species, figs are presented at different heights above ground level and at different locations (e.g. at ground-level runners, on stems or trunks, or in leaf axis; Berg & Wiebes 1992). The architecture of fruit display is likely to determine fruit suitability for particular frugivores, especially if frugivores exhibit different feeding behaviours. The variability of fruit presentation within *Ficus* thus allows discrete guilds of *Ficus* species to attract different subsets of the total frugivore community (Shanahan & Compton 2001). This might result in a distinct vertical stratification of fig-frugivore communities (the 'vertical stratification hypothesis'; Shanahan & Compton 2001) and could, at least partly, explain the positive relationship between *Ficus* and frugivore species richness.

All mechanisms outlined so far explain the positive relationship between food plant and animal consumer species richness with an increased availability of niches provided by a larger number of plant species (niche assembly hypothesis). In contrast, the species richness of trophically similar species (e.g. frugivores) competing for similar resources (e.g. fruits) could also result from stochastic ecological and evolutionary processes (Hubbell 2001). For instance, areas with high species richness of fleshy fruited plants could potentially produce more fruit biomass due to either more food plant individuals or higher total fruit production (e.g. Ortiz-Pulido & Rico-Gray 2000). If the total abundance of fruit resources increases with food plant species

richness, then more individuals of frugivores could be sustained in areas with high food plant diversity. The high species richness of frugivorous birds could then be governed by neutral speciation and extinction processes, where differences in traits of food plant and animal consumer species might be irrelevant for structuring plant-frugivore assemblages (Hubbell 2001; see also Burns 2006). In our case, *Ficus* species richness would then be positively associated with species richness of frugivorous birds, because it also correlates with the overall abundance and availability of fruit resources ('resource-abundance hypothesis').

Finally, the spatial congruence in patterns of fig and frugivore richness could not only be driven by figs as resources for frugivores, but also vice versa if frugivores constrain the spatial distribution and species richness of figs at continental scales. We tested this idea by interchanging *Ficus* richness and obligate frugivore richness in our path model (figure 2b) and found an influence of similar magnitude between frugivores and figs (direct effect, 0.532). This pattern could result if the large-scale distribution of fig species and their colonization of new sites are constrained by seed dispersal of frugivorous birds (e.g. Shanahan *et al.* 2001b for fig colonization of new volcanic islands). With a greater species richness of frugivorous dispersers, the seeds are more likely to arrive in a greater variety of sites and at different distances, as different species of birds have different foraging behaviours, perching locations and movement patterns. Furthermore, a higher species richness of frugivores might lead to better seed dispersal, more long-distance dispersal events and the foundation of new *Ficus* populations, potentially resulting in higher speciation rates of *Ficus* species.

In contrast to our study, many thoroughly conducted studies on plant-frugivore interactions have failed to document strong adaptive relationships (e.g. through demographic sorting or coevolutionary processes) between fruits and frugivores (e.g. Herrera 1998), suggesting that non-adaptive processes such as climate, historical or phylogenetic effects constrain the development of mutual adaptations (Herrera 2002). For instance, similar to our study, Márquez *et al.* (2004) analysed plant-frugivore richness at the scale of major river basins across Europe and found that avian frugivore richness was more dependent on environmental factors than on fleshy fruited plant species richness. Fleming (2005) examined the relationship between species richness of fruit-eating birds and their food plants in New and Old World communities and found hemispheric differences in plant-frugivore mutualisms. Recent plant-frugivore research suggests that these kinds of differences are often generated by analyses at different spatio-temporal scales (Burns 2004; García & Ortiz-Pulido 2004).

To understand the causal mechanisms of animal species richness patterns at continental and global scales, predictions from competing mechanistic hypotheses should be tested (Willig *et al.* 2003; Currie *et al.* 2004; Rahbek *et al.* 2007), ideally across multiple spatial and temporal scales (Böhning-Gaese 1997; Burns 2004; Rahbek 2005). Our results demonstrate a close relationship between the species richness of *Ficus* and avian frugivores in sub-Saharan Africa, suggesting that figs are keystone resources for animal consumers at continental scales. This relationship might

be driven by niche assembly mechanisms, e.g. coevolutionary adaptations to fruit size, fruit colour or vertical stratification of fruit presentation, or, alternatively, by a neutral speciation–extinction process. In both cases, however, the present study suggests that climatic variables influence frugivore species richness only indirectly via food webs rather than having a direct effect on the physiological tolerances of the organisms.

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1 APPENDIX S1: CLASSIFICATION OF FRUGIVORES

2 **Frugivore classification**

3 We used all 1,771 sub-Saharan breeding bird species in our database to classify them into
4 food guilds. The classification was based on food preference of each species as given in *The*
5 *Birds of Africa* (Brown *et al.* 1982; Urban *et al.* 1986; Fry *et al.* 1988; Keith *et al.* 1992;
6 Urban *et al.* 1997; Fry *et al.* 2000; Fry *et al.* 2004) and in the *Handbook of the Birds of the*
7 *World* (del Hoyo *et al.* 1992, 1994, 1996, 1997, 1999, 2001, 2002, 2003, 2004, 2005). Diet
8 categories were algae, amphibians and reptiles, aquatic invertebrates (aquatic insects and
9 crustaceans), birds, carrion, fish, fruit, mammals (e.g. rodents, bats, squirrels, monkeys),
10 nectar, seeds, omnivore, terrestrial plant parts (e.g. leaves, shoots, roots, flowers, bulbs),
11 terrestrial invertebrates (incl. spiders, insects, and mollusks), and other food items (e.g. wax,
12 human scraps, refuse). We distinguished major and minor food items by using keywords in
13 the paragraphs on food and feeding behavior (e.g. "almost exclusively", "entirely", "almost
14 entirely", "mainly", "prefers" taken to indicate major food items, and "occasionally",
15 "probably", "sometimes", "when available" etc. identifying minor food items). We then
16 classified three avian frugivore guilds depending on diet preference for fruits: (i) obligate
17 frugivores (the only major food item are fruits), (ii) partial frugivores (other major food items
18 besides fruits, e.g., terrestrial invertebrates), and (iii) opportunistic fruit-eaters (fruits only as
19 minor food items). The full list of frugivorous bird species is given in Appendix S2.

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55 APPENDIX S2: FRUGIVORE SPECIES LISTS

56 **List of obligate frugivores**

57 *Andropadus chlorigula* (inc. *fusc.*), *Andropadus gracilirostris*, *Andropadus importunes*, *Andropadus montanus*,
58 *Andropadus neumanni*, *Andropadus nigriceps*, *Andropadus tephrolaemus*, *Baeopogon clamans*, *Baeopogon*
59 *indicator*, *Buccanodon duchaillui*, *Ceratogymna albotibialis*, *Ceratogymna atrata*, *Ceratogymna brevis*,
60 *Ceratogymna bucinator*, *Ceratogymna cylindricus*, *Ceratogymna elata*, *Ceratogymna fistulator*, *Ceratogymna*
61 *subcylindricus*, *Chlorocichla falkensteini*, *Chlorocichla flavicollis*, *Chlorocichla simplex*, *Cinnyricinclus*
62 *femoralis*, *Cinnyricinclus leucogaster*, *Cinnyricinclus sharpii*, *Colius castanotus*, *Colius colius*, *Colius striatus*,
63 *Columba delegorguei*, *Columba iriditorques*, *Corythaeola cristata*, *Corythaixoides concolor*, *Corythaixoides*
64 *personatus*, *Crinifer piscator*, *Crinifer zonurus*, *Grafisia torquata*, *Gypohierax angolensis*, *Ixonotus guttatus*,
65 *Lamprotornis acuticaudus*, *Lamprotornis purpureiceps*, *Lioptilus nigricapillus*, *Lybius chaplini*, *Lybius dubius*,
66 *Lybius leucocephalus*, *Lybius rolleti*, *Lybius torquatus*, *Lybius vieilloti*, *Musophaga johnstoni*, *Musophaga*
67 *porphyreolopha*, *Musophaga rossae*, *Musophaga violacea*, *Onychognathus albirostris*, *Onychognathus blythii*,
68 *Onychognathus fulgidus*, *Onychognathus neumanni*, *Onychognathus tenuirostris*, *Parophasma galinieri*,
69 *Petronia pyrgita*, *Ploceus weynsi*, *Poeoptera kenricki*, *Poeoptera stuhlmanni*, *Pogoniulus atroflavus*, *Pogoniulus*
70 *bilineatus*, *Pogoniulus chrysoconus*, *Pogoniulus coryphaeus*, *Pogoniulus leucomystax*, *Pogoniulus pusillus*,
71 *Pogoniulus simplex*, *Pogoniulus subsulphureus*, *Pycnonotus barbatus*, *Pycnonotus dodsoni*, *Pycnonotus*
72 *somaliensis*, *Pycnonotus tricolor*, *Tauraco bannermani*, *Tauraco corythaix*, *Tauraco erythrolophus*, *Tauraco*
73 *fischeri*, *Tauraco hartlaubi*, *Tauraco leucolophus*, *Tauraco leucotis*, *Tauraco livingstonii*, *Tauraco*
74 *macrorhynchus*, *Tauraco persa*, *Tauraco ruspolii*, *Tauraco schalowi*, *Tauraco schuetti*, *Treron calva*, *Treron*
75 *waalia*, *Tricholaema diademata*, *Tricholaema hirsuta*, *Tricholaema lacrymosa*, *Urocolius indicus*, *Urocolius*
76 *macrourus*

77 **List of partial frugivores**

78 *Acryllium vulturinum*, *Afropavo congensis*, *Agapornis pullarius*, *Agapornis swindernianus*, *Agapornis taranta*,
79 *Amblyospiza albifrons*, *Anaplectes rubriceps*, *Andropadus ansorgei*, *Andropadus curvirostris*, *Andropadus*
80 *gracilis*, *Andropadus kakamegae*, *Andropadus latirostris*, *Andropadus masukuensis*, *Andropadus milanjensis*,
81 *Andropadus olivaceiceps*, *Andropadus virens*, *Anthoscopus flavifrons*, *Anthoscopus minutus*, *Anthreptes*
82 *anchietae*, *Anthreptes aurantium*, *Anthreptes axillaris*, *Anthreptes collaris*, *Anthreptes fraseri*, *Anthreptes*
83 *rectirostris*, *Bubalornis albirostris*, *Calyptocichla serina*, *Cercomela sinuata*, *Chlorocichla flaviventris*,
84 *Chlorocichla laetissima*, *Chlorocichla prigoginei*, *Cichladusa ruficauda*, *Coccycolius iris*, *Colius leucocephalus*,

85 *Columba albinucha, Columba arquatrix, Columba larvata, Columba sjostedti, Columba uncinata,*
86 *Corythaixoides leucogaster, Cossypha caffra, Cossypha humeralis, Cossypha natalensis, Cossypha niveicapilla,*
87 *Cossypha roberti, Creatophora cinerea, Criniger barbatus, Dendrocygna bicolor, Estrilda astrild, Estrilda*
88 *caerulescens, Francolinus adpersus, Francolinus afer, Francolinus achantensis, Francolinus bicalcaratus,*
89 *Francolinus camerunensis, Francolinus capensis, Francolinus clappertoni, Francolinus erckelii, Francolinus*
90 *hartlaubi, Francolinus harwoodi, Francolinus icterorhynchus, Francolinus jacksoni, Francolinus leucoscepus,*
91 *Francolinus levaillantoides, Francolinus natalensis, Francolinus ochropectus, Francolinus squamatus,*
92 *Francolinus swainsonii, Guttera pucherani, Gymnobucco bonapartei, Gymnobucco calvus, Gymnobucco peli,*
93 *Gymnobucco sladeni, Histurgops ruficauda, Lamprotornis caudatus, Lamprotornis chalcurus, Lamprotornis*
94 *chalybaeus, Lamprotornis chloropterus, Lamprotornis corruscus, Lamprotornis cupreocauda, Lamprotornis*
95 *elisabeth, Lamprotornis mevesii, Lamprotornis nitens, Lamprotornis pulcher, Lamprotornis purpureus,*
96 *Lamprotornis purpuropterus, Lamprotornis shelleyi, Lamprotornis splendidus, Laniarius bicolour, Laniarius*
97 *erythrogaster, Lybius bidentatus, Lybius guifsobalito, Lybius melanopterus, Lybius minor, Lybius rubrifacies,*
98 *Lybius undatus, Malimbus ibadanensis, Melignomon eisentrauti, Monticola rupestris, Nectarinia johanna,*
99 *Nectarinia olivacea, Nectarinia pembae, Nectarinia rubescens, Nectarinia violacea, Nigrita bicolor, Nigrita*
100 *canicapilla, Nigrita fusconota, Nigrita luteifrons, Onychognathus morio, Onychognathus nabouroup,*
101 *Onychognathus salvadorii, Onychognathus walleri, Oriolus auratus, Oriolus chlorocephalus, Oriolus larvatus,*
102 *Oriolus nigripennis, Oriolus percivali, Phyllastrephus strepitans, Ploceus albinucha, Ploceus aurantius, Ploceus*
103 *aureonucha, Ploceus bicolor, Ploceus cucullatus, Ploceus golandi, Ploceus insignis, Ploceus tricolor, Ploceus*
104 *velatus, Ploceus xanthops, Poeoptera lugubris, Pogoniulus scolopaceus, Pogonocichla stellata, Poicephalus*
105 *cryptoxanthus, Poicephalus flavifrons, Poicephalus gulielmi, Poicephalus meyeri, Poicephalus robustus,*
106 *Poicephalus rueppellii, Poicephalus rufiventris, Poicephalus senegalus, Polyboroides typus, Pseudocalyptomena*
107 *graueri, Psittacula krameri, Psittacus erithacus, Ptilopachus petrosus, Ptilostomus afer, Pycnonotus capensis,*
108 *Pycnonotus nigricans, Rhynchostruthus socotranus, Serinus albogularis, Serinus burtoni, Serinus canicapillus,*
109 *Serinus citrinelloides, Serinus gularis, Serinus leucopterus, Serinus mennelli, Serinus scotops, Serinus*
110 *sulphuratus, Serinus whytii, Sheppardia gunningi, Speculipastor bicolour, Speirops melanocephalus,*
111 *Spermophaga haematina, Spreo albicapillus, Stactolaema anchietae, Stactolaema leucotis, Stactolaema*
112 *olivacea, Stactolaema whytii, Sylvia boehmi, Sylvia layardi, Sylvia leucomelaena, Sylvia subcaeruleum,*
113 *Thamnolaea cinnamomeiventris, Thamnolaea coronata, Thescelocichla leucopleura, Tockus alboterminatus,*
114 *Tockus fasciatus, Tockus flavirostris, Trachyphonus darnaudii, Trachyphonus erythrocephalus, Trachyphonus*
115 *margaritatus, Trachyphonus purpuratus, Trachyphonus usambiro, Trachyphonus vaillantii, Tricholaema*

116 *frontata*, *Tricholaema leucomelas*, *Tricholaema melanocephala*, *Turdoides fulvus*, *Turdus helleri*, *Turdus*
117 *olivaceus*, *Turdus pelios*, *Turdus roehli*, *Turdus smithii*, *Turtur tympanistria*, *Zoothera gurneyi*, *Zoothera*
118 *piaggiae*, *Zoothera tanganjicae*, *Zosterops kulalensis*, *Zosterops pallidus*, *Zosterops poliogaster*, *Zosterops*
119 *silvanus*, *Zosterops winifredae*

120 **List of opportunistic fruit-eaters**

121 *Agapornis fischeri*, *Agapornis lilianae*, *Agelastes meleagrides*, *Agelastes niger*, *Alethe fuelleborni*, *Anas*
122 *hottentota*, *Anthoscopus caroli*, *Anthoscopus sylviella*, *Anthreptes longuemarei*, *Anthreptes rubritorques*, *Apalis*
123 *flavida*, *Apalis thoracica*, *Ardeotis arabs*, *Ardeotis kori*, *Bleda canicapillus*, *Bleda eximinius*, *Bleda notatus*,
124 *Bleda syndactylus*, *Bradornis mariquensis*, *Bradornis pallidus*, *Bubalornis niger*, *Bubo leucostictus*, *Bubo*
125 *poensis*, *Bubo vosseleri*, *Bubulcus ibis*, *Bucorvus abyssinicus*, *Bucorvus cafer*, *Camaroptera brachyura*,
126 *Camaroptera brevicaudata*, *Camaroptera harterti*, *Campephaga flava*, *Cercomela familiaris*, *Cercomela*
127 *melanura*, *Cercotrichas coryphaeus*, *Cercotrichas galactotes*, *Cercotrichas leucophrys*, *Cercotrichas paena*,
128 *Cercotrichas signata*, *Certhilauda albescens*, *Certhilauda benguelensis*, *Certhilauda brevirostris*, *Certhilauda*
129 *burra*, *Certhilauda curvirostris*, *Certhilauda semitorquata*, *Certhilauda subcoronata*, *Ceuthmochares aereus*,
130 *Chrysococcyx cupreus*, *Chrysococcyx flavigularis*, *Chrysococcyx klaas*, *Cichladusa guttata*, *Columba guinea*,
131 *Columba livia*, *Columba oliviae*, *Coracias cyanogaster*, *Coracina azurea*, *Coracina caesia*, *Corvinella*
132 *melanoleuca*, *Corvus capensis*, *Corvus crassirostris*, *Corvus rhipidurus*, *Cosmopsarus regius*, *Cosmopsarus*
133 *unicolor*, *Cossypha cyanocampter*, *Cossypha dichroa*, *Cossypha heuglini*, *Criniger calurus*, *Criniger*
134 *chloronotus*, *Criniger ndussumensis*, *Criniger olivaceus*, *Cuculus solitarius*, *Dendrocycna viduata*, *Dendropicus*
135 *fuscescens*, *Dendropicus obsoletus*, *Dicrurus adsimilis*, *Dinemellia dinemelli*, *Dioptrornis fischeri*, *Dryoscopus*
136 *cubla*, *Eremomela badiceps*, *Eremomela pusilla*, *Eremopterix australis*, *Euplectes ardens*, *Euplectes hartlaubi*,
137 *Eupodotis gindiana*, *Eupodotis melanogaster*, *Eupodotis ruficrista*, *Eupodotis savilei*, *Eupodotis senegalensis*,
138 *Eupodotis vigorsii*, *Eurocephalus anguitimens*, *Eurocephalus rueppelli*, *Eurystomus gularis*, *Falco ardosiaceus*,
139 *Francolinus africanus*, *Francolinus lathamii*, *Francolinus sephaena*, *Fraseria cinerascens*, *Fraseria ocreata*,
140 *Fulica cristata*, *Galerida magnirostris*, *Gallinula chloropus*, *Guttera plumifera*, *Halcyon malimbica*, *Hippolais*
141 *pallida*, *Hirundo abyssinica*, *Hirundo cucullata*, *Hyliota violacea*, *Illadopsis pyrrhoptera*, *Indicator conirostris*,
142 *Indicator exilis*, *Indicator indicator*, *Indicator maculatus*, *Indicator meliphilus*, *Indicator variegatus*, *Kupeornis*
143 *rufocinctus*, *Lamprotornis australis*, *Lamprotornis hildebrandti*, *Lamprotornis superbus*, *Laniarius*
144 *atrococcineus*, *Laniarius ferrugineus*, *Laniarius funebris*, *Laniarius mufumbiri*, *Lanius cabanisi*, *Lanius collaris*,
145 *Larus leucophthalmus*, *Linurgus olivaceus*, *Lonchura bicolor*, *Lonchura nigriceps*, *Lophaetus occipitalis*,
146 *Macrosphenus flavicans*, *Malcorus pectoralis*, *Malimbus cassini*, *Malimbus erythrogaster*, *Malimbus*

147 *malimbicus*, *Malimbus nitens*, *Malimbus rubricollis*, *Mandingoa nitidula*, *Melaenornis pammelaina*,
148 *Melichneutes robustus*, *Modulatrix stictigula*, *Monticola brevipes*, *Monticola explorator*, *Monticola pretoriae*,
149 *Monticola rufocinereus*, *Muscicapa adusta*, *Muscicapa caerulescens*, *Muscicapa comitata*, *Muscicapa epulata*,
150 *Muscicapa infuscata*, *Myioparus griseigularis*, *Myrmecocichla aethiops*, *Myrmecocichla formicivora*,
151 *Myrmecocichla nigra*, *Namibornis herero*, *Nectarinia afra*, *Nectarinia batesi*, *Nectarinia chalybea*, *Nectarinia*
152 *chloropygia*, *Nectarinia cyanolaema*, *Nectarinia hunteri*, *Nectarinia osea*, *Nectarinia seimundi*, *Nectarinia*
153 *superba*, *Nectarinia ursulae*, *Neocossyphus fraseri*, *Neocossyphus poensis*, *Neolestes torquatus*, *Neotis denhami*,
154 *Neotis heuglinii*, *Neotis nuba*, *Nesocharis capistrata*, *Nesocharis shelleyi*, *Nicator chloris*, *Numida meleagris*,
155 *Oenanthe leucopyga*, *Oenanthe monticola*, *Oriolus brachyrhynchus*, *Oriolus monacha*, *Oxylophus jacobinus*,
156 *Oxylophus levaillantii*, *Pachyphantes superciliosus*, *Parmoptila woodhousei*, *Parus leucomelas*, *Parus niger*,
157 *Passer diffusus*, *Passer griseus*, *Passer luteus*, *Passer melanurus*, *Philetairus socius*, *Phoeniculus bollei*,
158 *Phoeniculus castaneiceps*, *Phoeniculus purpureus*, *Phragmacia substriata*, *Phyllanthus atripennis*,
159 *Phyllastrephus albigularis*, *Phyllastrephus alfredi*, *Phyllastrephus baumanni*, *Phyllastrephus cabanisi*,
160 *Phyllastrephus cerviniventris*, *Phyllastrephus flavostriatus*, *Phyllastrephus hypochloris*, *Phyllastrephus*
161 *icterinus*, *Phyllastrephus placidus*, *Phyllastrephus poensis*, *Phyllastrephus scandens*, *Phyllastrephus terrestris*,
162 *Phyllastrephus xavieri*, *Platysteira castanea*, *Platysteira concreta*, *Plectropterus gambensis*, *Ploceus alienus*,
163 *Ploceus baglafecht*, *Ploceus capensis*, *Ploceus heuglini*, *Ploceus intermedius*, *Ploceus melanogaster*, *Ploceus*
164 *nigerrimus*, *Ploceus nigricollis*, *Ploceus ocularis*, *Ploceus preussi*, *Porphyrio alleni*, *Porphyrio porphyrio*,
165 *Prinia maculosa*, *Prionops caniceps*, *Prionops plumatus*, *Prionops rufiventris*, *Prionops scopifrons*, *Prodotiscus*
166 *insignis*, *Prodotiscus regulus*, *Prodotiscus zambesiae*, *Pseudoalcippe abyssinica*, *Pseudoalcippe atriceps*,
167 *Psophocichla litsipsirupa*, *Pyrenestes ostrinus*, *Pyrrhocorax pyrrhocorax*, *Rhinopomastus aterrimus*,
168 *Rhinopomastus cyanomelas*, *Rhinopomastus minor*, *Rhodophoneus cruentus*, *Saxicola torquata*, *Serinus alario*,
169 *Serinus ankoberensis*, *Serinus atrogularis*, *Serinus frontalis*, *Serinus leucolaema*, *Serinus mozambicus*, *Serinus*
170 *striolatus*, *Sigelus silens*, *Sphenoaecus afer*, *Spreo bicolor*, *Spreo fischeri*, *Streptopelia capicola*, *Streptopelia*
171 *decipiens*, *Streptopelia lugens*, *Streptopelia reichenowi*, *Streptopelia semitorquata*, *Streptopelia senegalensis*,
172 *Streptopelia vinacea*, *Struthio camelus*, *Swynnertonia swynnertoni*, *Sylvia lugens*, *Sylvia rueppelli*, *Sylvietta*
173 *virens*, *Tchagra senegala*, *Tchagra tchagra*, *Telophorus bocagei*, *Telophorus olivaceus*, *Telophorus zeylonus*,
174 *Terpsiphone rufiventer*, *Terpsiphone viridis*, *Tockus albocristatus*, *Tockus bradfieldi*, *Tockus camurus*, *Tockus*
175 *deckeni*, *Tockus erythrorhynchus*, *Tockus hartlaubi*, *Tockus hemprichii*, *Tockus jacksonii*, *Tockus leucomelas*,
176 *Tockus monteiri*, *Tockus nasutus*, *Turdoides jardineii*, *Turdoides melanops*, *Turdoides reinwardtii*, *Turdoides*

- 177 *sharpei*, *Turdoides tenebrosus*, *Turdus libonyanus*, *Turdus tephronotus*, *Uraeginthus granatina*, *Zoothera*
178 *guttata*, *Zosterops senegalensis*

180 **List of fig species**

- 181 *Ficus abscondita*, *Ficus abutilifolia*, *Ficus adolfi-friderici*, *Ficus amadiensis*, *Ficus ardisioides*, *Ficus*
182 *artocarpoides*, *Ficus asperifolia*, *Ficus barteri*, *Ficus bizanae*, *Ficus bubu*, *Ficus burretiana*, *Ficus burtt-davyi*,
183 *Ficus bussei*, *Ficus calyptrata*, *Ficus capreifolia*, *Ficus chirindensis*, *Ficus chlamydocarpa*, *Ficus conraui*, *Ficus*
184 *cordata*, *Ficus crassicosta*, *Ficus craterostoma*, *Ficus cyathistipula*, *Ficus cyathistipuloides*, *Ficus*
185 *densistipulata*, *Ficus dicranostyla*, *Ficus dryepontiana*, *Ficus elasticoides*, *Ficus exasperata*, *Ficus*
186 *faulkneriana*, *Ficus fischeri*, *Ficus glumosa*, *Ficus ilicina*, *Ficus ingens*, *Ficus jansii*, *Ficus kamerunensis*, *Ficus*
187 *leonensis*, *Ficus lingua*, *Ficus louisii*, *Ficus lutea*, *Ficus lyrata*, *Ficus modesta*, *Ficus mucoso*, *Ficus*
188 *muelleriana*, *Ficus natalensis*, *Ficus nigropunctata*, *Ficus oreodryadum*, *Ficus oresbia*, *Ficus ottoniifolia*, *Ficus*
189 *ovata*, *Ficus pachyneura*, *Ficus palmata*, *Ficus persicifolia*, *Ficus platyphylla*, *Ficus polita*, *Ficus populifolia*,
190 *Ficus preussii*, *Ficus pseudomangifera*, *Ficus psilopoga*, *Ficus pygmaea*, *Ficus recurvata*, *Ficus rokko*, *Ficus*
191 *sagittifolia*, *Ficus salicifolia*, *Ficus sansibarica*, *Ficus saussureana*, *Ficus scasselatii*, *Ficus scott-elliottii*, *Ficus*
192 *stuhmannii*, *Ficus subcostata*, *Ficus subsagittifolia*, *Ficus sur*, *Ficus sycomorus*, *Ficus tesselata*, *Ficus tettensis*,
193 *Ficus thonningii*, *Ficus tremula*, *Ficus trichopoda*, *Ficus umbellata*, *Ficus usambarensis*, *Ficus vallis-choudae*,
194 *Ficus variifolia*, *Ficus vasta*, *Ficus verruculosa*, *Ficus vogeliana*, *Ficus wakefieldii*, *Ficus wildemaniana*

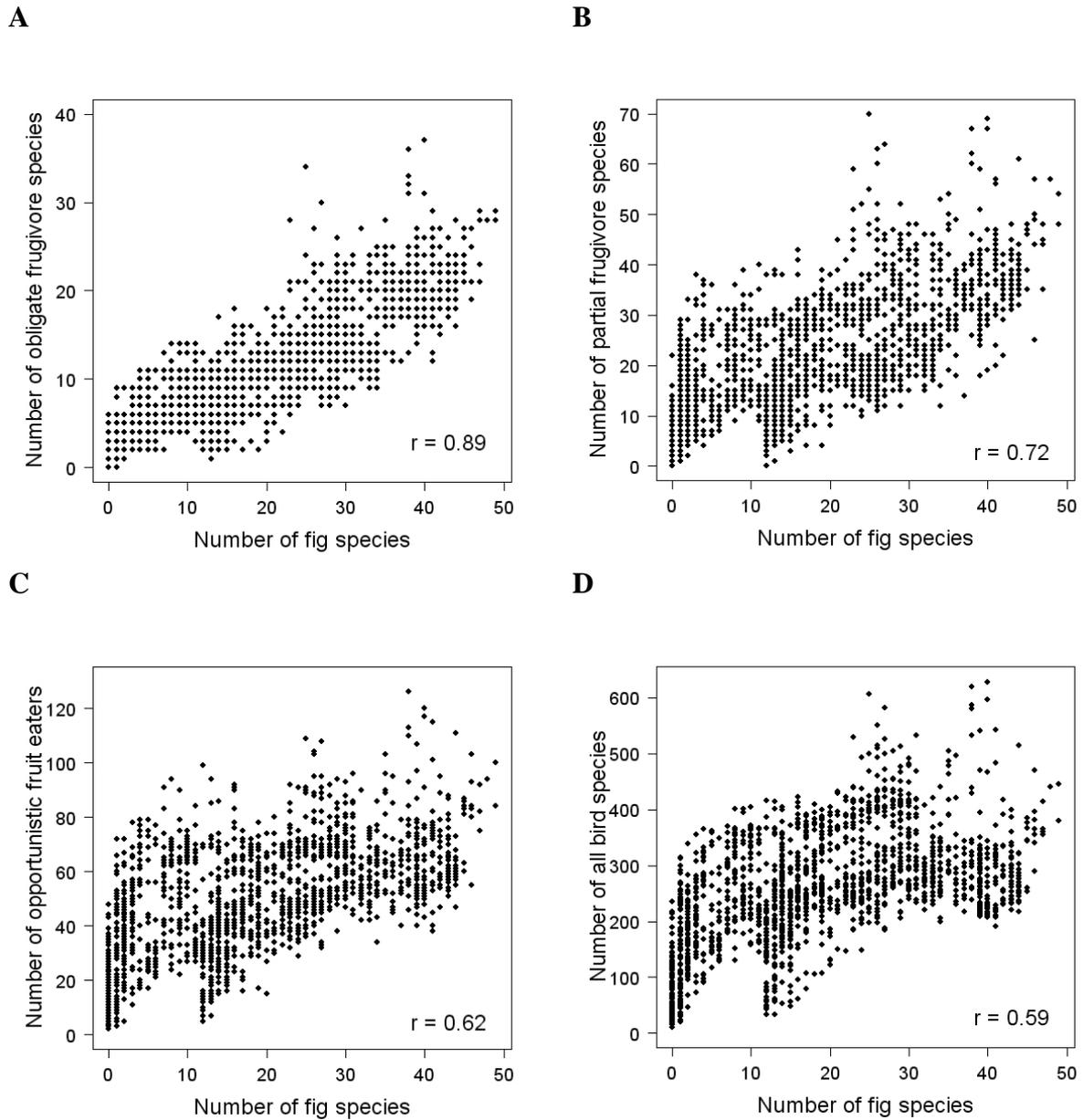


Figure S4. The relationship between fig (*Ficus* spp.) richness and species richness of obligate frugivores (A), partial frugivores (B), opportunistic fruit-eaters (C), and all breeding birds (D) in sub-Saharan Africa. Spearman rank correlations are given in the lower right corner of each graph.

197 APPENDIX TABLE S5: CORRELATION MATRIX

198 Table S5. Correlation matrix of untransformed predictor and response variables. OBL = obligate frugivores ($n =$
 199 92); PAR = partial frugivores ($n = 200$); OPP = opportunistic frugivores ($n = 290$); ALL = all bird species ($n =$
 200 1,772). Mnemonics of predictor variables are explained in Table 1.

predictor variables	spearman rank correlation (r_s)									
	bird richness				predictor variables					
	OBL	PAR	OPP	ALL	FigRich	Prec	MaxTemp	NPP	AltRange	EcoDiv
FigRich	0.89	0.72	0.62	0.59	1					
Prec	0.87	0.71	0.63	0.60	0.89	1				
MaxTemp	-0.33	-0.59	-0.65	-0.58	-0.20	-0.29	1			
NPP	0.85	0.75	0.68	0.63	0.85	0.93	-0.36	1		
AltRange	0.19	0.34	0.37	0.41	0.10	0.06	-0.52	0.06	1	
EcoDiv	0.57	0.68	0.66	0.67	0.46	0.48	0.45	0.51	0.34	1

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